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Sarcoptic Mange Research in Black Bears

For the last six years, SCWDS has been involved in collaborative research to understand more about the emergence and expansion of sarcoptic mange in black bears (Ursus americanus) in the northeastern and mid-Atlantic states (https://doi.org/10.1016/j.vprsr.2019.100303). Δ former Master's student at SCWDS, Sarah Knox Peltier, provided foundational research exploring genetic variation of the parasites in bears and showed that this emergence was not the result of a single, novel, highly-pathogenic mite strain (https://doi.org/10.1645/17-26). Additionally, her research showed that skin scraping was the diagnostic test that most likely resulted in a diagnosis of mange and the ability to identify the causative mite as compared to other more costly labor-intensive tests and (https://doi.org/10.7589/2017-06-148). More recently. doctoral student, а Dr. Kevin Niedringhaus, continued this work by exploring the ability of mites to survive off of the host, whether there are associations between sarcoptic mange and evidence of prior exposure to pathogens commonly present in bears, and if there is widespread exposure of bears to sarcoptic mange mites across the landscape. Results from Dr. Niedringhaus' recently completed research are highlighted below.

Our first objective was to estimate the extent of presumed exposure of bears to *S. scabiei* throughout Pennsylvania, the state currently experiencing the highest number of mange cases diagnosed in bears. There was anecdotal evidence that bears in Northeastern Pennsylvania, particularly in the Poconos, were relatively free of the disease while other parts of the state, namely the central area, were documented to have many cases of mange. Serology, a tool that indirectly measures whether an animal has been exposed to a pathogen by detecting circulating antibodies specific to that pathogen, can provide valuable information on the epidemiology of mange in bears in these regions. If antibodies to mites were widespread in both regions, it would suggest that bears in the central part of the state were predisposed, by some unknown factor, to developing disease associated with *S. scabiei* infection. However, if the overall antibody prevalence was low, we could speculate that all bears are inherently susceptible to developing mange after coming into contact with mites and that the presence of specific risk factors in some populations may be less likely explanations of the apparent variation in disease distribution.

Before research to explore these concepts could begin, a serological test had to be validated because no test was commercially developed for use in black bears. We evaluated a test designed to detect antibodies against S. scabiei in dogs. Initially, samples from bears with confirmed sarcoptic mange were used in conjunction with samples from cubs born to seronegative sows to ensure that the test could reliably detect antibodies in known positive cases and were reliably negative in presumed non-exposed cases. Additionally, serum samples collected sequentially from bears after they had received treatment for sarcoptic mange were supplied by collaborators at the Wildlife Center of Virginia to determine the extent that antibodies can still be detected after the disease resolves. This is an important concept because if antibodies were quickly depleted, we would expect our field prevalence estimates to be artificially low.

After confirming that the test was accurate to an acceptable level and understanding the persistence of antibodies over time, 437 samples were collected from harvested bears at check stations in Pennsylvania over two years. These

samples were divided into two groups: samples from bears originating in high-mange areas and samples from bears in low-mange areas. The seroprevalence of non-diseased bears in the highand low-mange area was 6.7% and 0%, respecitvely. These data not only show that there is presumably a lack of exposure to mites in the low-mange area but also that few bears in highmange areas are being exposed without developing clinical disease.



Figure A: Magnified view of crusts on the skin of a bear with severe sarcoptic mange showing an embedded mite (arrow).

Figure B: High-magnification view of an adult female S. scabiei as seen from a skin scrape from a bear with severe mange.

These data provide evidence that bears are inherently susceptible to disease after exposure. This finding has important implications for bear populations in other regions, as there is evidence of the continued disease expansion, including recent confirmed cases in Oklahoma and Arkansas. Additionally, the validation of the serological assay can be used by SCWDS and others to continue research on the epidemiology of mange in bears.

The research that SCWDS students conducted on this subject, as well as a general overview of mange in wildlife, was recently presented at the Eastern Black Bear Workshop in Potosi, Missouri, on April 23, 2019. The presentation was designed to inform biologists and other agency personnel of our current knowledge of the disease, theories on its emergence, and potential implications for bears in the future. Also, for those interested in learning more about sarcoptic mange, we recently published an open access peer-reviewed review of the disease in wildlife in North America (<u>https://doi.org/10.1016/j.ijppaw.2019.06.003</u>). (Prepared by Kevin Niedringhaus and Michael

(Prepared by Kevin Niedringhaus and Michael Yabsley)

Research into the Effects of West Nile Virus on Gamebirds

Since its introduction to the United States in 1999. West Nile virus (WNV) has spread rapidly and is now considered endemic throughout much of North America. WNV is a vector-borne flavivirus that is most commonly transmitted by mosquitoes in the Culex genus and circulates between mosquitoes, birds, and mammalian hosts (including humans). Numerous bird species can be infected with WNV but clinical outcome can vary significantly between species, ranging from inapparent infection to death. This varied clinical spectrum appears to occur between galliform species. For example, results from early WNV research indicate that some domesticated galliforms (e.g., chickens and domestic turkeys) are resistant to WNV-associated disease. In contrast, greater sage grouse (Centrocercus urophasianus) and more recently, ruffed grouse (Bonasa umbellus), are highly susceptible to WNVassociated morbidity and mortality. The potential impacts of WNV on other upland game bird species such as wild turkey (*Meleagris gallopavo silvestris*) and bobwhite quail (Colinus virginianus) are largely unknown.

Recently, SCWDS researchers have begun collaborating with wildlife biologists and veterinarians from multiple state agencies across the eastern United States as well as central and eastern Canada to test hunter-harvested, ruffed grouse blood samples for antibodies to WNV. These findings will ultimately be used to develop a multi-year, WNV antibody prevalence data set on ruffed grouse populations across part of their range in eastern North America. The detection of antibodies to WNV in the blood of wild birds generally indicates either prior WNV infection (most likely via mosquito bite) or maternallyderived antibodies (antibodies passed from mother to egg and chick that wane over time). This project developed in response to concerns among numerous state wildlife agencies over ruffed grouse population declines and the current research supporting the potential for WNV to impact ruffed grouse health.

In response to similar concerns regarding depressed population numbers and low recruitment rates in eastern wild turkey and northern bobwhite quail in some areas, SCWDS researchers are also exploring the potential impact of WNV on these species. During the summer and fall of 2019, eastern wild turkey poults and juvenile northern bobwhite quail will be experimentally infected with WNV. Groups of turkey and quail will be inoculated with WNV at 6 and 16 weeks posthatch and monitored for two weeks following inoculation for signs of clinical disease, viral shedding (oropharyngeal and cloacal), and virus in blood (viremia). Postmortem evaluations will assess for pathology and evidence of virus persistence in tissues. Results of this research will provide critical insight into the potential for WNV to impact these upland game bird species and will be useful in interpreting field data. State agencies will be able to consider these data in future management strategies of upland game birds.

We thank the Multistate Conservation Grant Program for funding this study as well as the Pennsylvania Game Commission (PGC) for providing the eastern wild turkey eggs. (Prepared by Melanie Kunkel and Nicole Nemeth)

Snow Geese Mortality Event in Kansas

During March 2019, the morning light revealed a grisly scene on a northwestern Kansas prairiewetland. Approximately 140 dead and dying snow and Ross's geese (Chen caerulescens and C. rossii, respectively) were observed near a pond in Norton County, Kansas (Figure 1). Ninety percent of the geese had blood discharging from the ears (Figure 2) and about 25% of these had obvious skull fractures, blood on the feathers of the head, and bulging eyes; others had severed heads. The night before, severe thunderstorms with lightning moved through the area, including strong wind gusts, up to 60 mph. The carcasses of five geese were submitted to SCWDS by the Kansas Department of Wildlife, Parks, and Tourism (KDWPT) for further examination.

Postmortem examination confirmed the field findings of acute skull fractures with hemorrhage, and also revealed blood in the oral cavity, eyes, and ear canals as well as acute spinal fractures with hemorrhage. Examination revealed that all birds were in good nutritional condition and had recently eaten based on digestive tract contents. Microscopic examination revealed hemorrhage in the lung of one bird, but otherwise normal tissues in all five geese. There was no evidence of an infectious process, including avian cholera (caused by *Pasteurella multocida*), which was ruled out by laboratory tests. Based primarily on the detailed history and field findings provided by KDWPT personnel, gross findings, and lack of other evidence of disease, these geese were diagnosed with severe, acute blunt-force trauma, primarily to the head, as the cause of death. Violent storms in the area the night prior to finding the geese was deemed the likely source of trauma, possibly through strong wind gusts or lightning strike during flight which may have caused confusion and disorientation of the flock leading to collision with each other and the ground. These sources cannot be confirmed, but the gross presentation suggests severe, bluntforce (and high velocity) head trauma, strong enough to cause decapitation in some cases.



Figure 1.

Figure 2.

Snow geese often travel in flocks of dozens of birds or more, which can be joined by smaller numbers of Ross's geese, especially during migration when they often fly at high altitude during the night. This makes them susceptible to large mortality events when they encounter large-scale trauma-inducing factors, toxins, or a highly contagious, infectious pathogen. In the present case, the number of affected birds was between 100 and 200, which were all found within relatively close proximity on the landscape. There have been anecdotal reports of single, dead birds falling from the sky due to lightning strike, which remains an alternative possibility as to the cause of death in these geese in Kansas. Birds may have fallen at high speeds and hit the ground, which could have caused the observed gross findings. Further, the high numbers of affected geese may arise due to confusion and disorientation within the night-flying flock following

the lightning storm. Strong and unpredictable wind gusts would also likely contribute.

Population numbers of both snow and Ross's geese across their range from Mexico north to the Arctic have steadily increased since the 1950s, believed to be facilitated by warming climates in their northern breeding grounds, allowing for increased access to the vegetation they eat, as well as a lengthened growing season. Concurrent eastward range expansion of Ross's geese has led to increased geographic overlap of the two species, resulting in hybridization. Both species are federally protected game bird species with annual hunting seasons that vary by region.

SCWDS would like to thank the KDWPT for submission of this case, and participation in the case summary and BRIEFS article. (Prepared by Nicole Nemeth, SCWDS; Shane Hesting, and Luke Winge, KDWPT.)

A Note to Our Readers

We thank you for your sustained interest in our quarterly newsletter, the SCWDS BRIEFS. We continue to receive positive feedback from many readers, which lets us know that we are still providing items of interest to you in each issue.

One difficult aspect of putting out a publication such as the BRIEFS is maintaining the mailing list. We want to reach as many of you as we can, but can do so only if you let us know you want to be included on the mailing list, notify us of any address changes, or inform us of someone else you know who would like to be added to the mailing list. Of course, if you want to reduce the volume of mail coming into your home or office, you may opt to be removed from the regular mailing list and have your name added to our email list to be informed when each new issue is posted on our website. This way, you usually can read the newsletter at least 10 days before a mailed copy would arrive. As always, if you have suggestions for improvement of the BRIEFS, please let us hear from you. Our goal is to provide information of interest to you.

Recent SCWDS Publications Available

Below are some recent publications authored or co-authored by SCWDS staff. Many of these can be accessed online from the web pages of the various journals. If you do not have access to this service and would like to have a copy of any of these papers, let us know. Many can be sent to you electronically with minimum effort; others will be mailed to you. For your convenience, please indicate requested publications, fill out the form on page 7, and check the appropriate box to receive either an electronic copy or a hard copy and return it to us: SCWDS, College of Veterinary Medicine, University of Georgia, Athens, GA 30602 or email at brewton@uga.edu.

- Allen, S.E., J.L. Rothenburger, C.M. Jardine, A. Ambagala, K. Hooper-McGrevy, N. Colucci, T. Furukawa-Stoffer, S. Vigil, M. Ruder, and N.M. Nemeth. 2019. Epizootic hemorrhagic disease in wild white-tailed deer, Canada. *Emerging Infectious Diseases*. 25(4):832-834.
- Astorga, F., S. Carver, E. Almberg, G.R. Sousa,
 K. Wingfield, K.D. Niedringhaus, P. Van Wick, L.
 Rossi, Y, Xie, P. Cross, S. Angelone, C. Gortazar,
 L.E. Escobar. 2018. Meeting Report:
 International meeting on sarcoptic mange in
 wildlife, June 2018, Blacksburg, Virginia, USA. *Parasites and Vectors* 11(1): 449.
- Beard, C.B., J. Occi, D.L. Bonilla, A.M. Egizi, D.M. Fonseca, J.W. Mertins, B.P. Backenson, W.I. Bajwa, A.M. Barbarin, M.A. Bertone, J. Brown, N.P. Connally, N.D. Connell, R.J. Eisen, R.C. Falco, A.M. James, R.K. Krell, K. Lahmers, N. Lewis, S.E. Little, M. Neault, A.A. Perez de Leon, A.R. Randall, M.G. Ruder, M.N. Saleh, B.L. Schappach, B.A. Schroeder, L.L. Seraphin, M. Wehtje, G.P. Wormser, M.J. Yabsley, and W. Halperin. 2018. Multistate infestation with the exotic disease-vector tick *Haemaphysalis longicornis* - United States, August 2017-September 2018. *Morbidity and Mortality Weekly Report* 67(47): 1310-1313.
- Becker, D.J., C.S. Teitelbaum, M.H. Murray, S.E. Curry, C.N. Welch, T. Ellison, H.C. Adams, R.S. Rozier, E.K. Lipp, S.M. Hernandez, S. Altizer, and R. Hall. 2018. Assessing the contributions of intraspecific and environmental sources of infection in urban wildlife: *Salmonella enterica* and white ibis as a case study. *Journal of the Royal Society Interface* 15(149): 20180654.
- Bloodgood, J.C., T.M. Norton, L.A. Hoopes, N.I. Stacy, and S.M. Hernandez. 2019. Comparison of hematological, plasma biochemical, and nutritional analytes of rehabilitating and apparently healthy free-ranging Atlantic green turtles (*Chelonia mydas*). *Journal of Zoo and Wildlife Medicine* 50(1): 69-81.

- Carlton, R.E., H. Fenton, J.A. Bryan, and M.J. Yabsley. 2019. Pathology in Practice: Xanthoma in a great horned owl. *Journal of the American Veterinary Medical Association* 254(12): 1407-1409.
- Carter, D., R. Poulson, P. Link, P. Walther, A. Ramey, and D.E. Stallknecht. 2018. Influenza A prevalence and subtype diversity in migrating teal sampled along the United States Gulf Coast. *Avian Diseases* 63 (Suppl) 165-171.
- Clarke, L.L., M.G. Ruder, C. Kienzle-Dean, D. Carter, D.E. Stallknecht, and E.W. Howerth. 2019. Experimental infection of white-tailed deer (*Odocoileus virginianus*) with bluetongue virus serotype 3. *Journal of Wildlife Diseases* doi: 10.7589/2018-06-159.
- Clarke, L.L., M.G. Ruder, D.G. Mead, and E.W. Howerth. 2018. Heartland virus exposure in white-tailed deer in the southeastern United States, 2001-2015. *American Journal of Tropical Medicine and Hygiene* 99(5): 1346-1349.
- Cleveland, C.A., M.L. Eberhard, A.T. Thompson, K.B. Garrett, L. Swanepoel, H. Zirimwabagabo, T. Moundai, P.T. Ouakou, E. Ruiz-Tiben, and M.J. Yabsley. 2019. A search for tiny dragons (*Dracunculus medinensis* third stage larvae) in aquatic animals, Chad, Africa. *Scientific Reports* 9(1): 375.
 - Cleveland, C.A., K.B. Garrett, R.A. Cozad, B.M. Williams, M.H. Murray, and M.J. Yabsley. 2018. The wild world of Guinea worms: A review of the genus *Dracunculus* in wildlife. *International Journal of Parasitology Parasites Wildlife* 7(3): 289-300.
- Cleveland, C.A., L. Swanepoel, E.K. Box, A. De Nicola, and M.J. Yabsley. 2019. Rickettsia species in ticks collected from wild pigs (*Sus scrofa*) and Philippine deer (*Rusa marianna*) on Guam, Marianna Islands, USA. Acta Tropica. 18(194): 89-92.
- Crum, J.A., D.G. Mead, M.W. Jackwood, J.E. Phillips, and D.E. Stallknecht. 2018. Genetic relatedness of epizootic hemorrhagic disease virus serotype 2 from the 2012 outbreak in the USA. *Journal of Wildlife Diseases* 55(2): 363-374.
 - Elsmo, E.J., and H. Fenton. 2019. Pituitary abscesses in four free-ranging white-tailed deer (*Odocoileus virginianus*). *Journal of Wildlife Diseases* 55(1): 254-257.
- Elsmo, E.J., H. Fenton, C.A. Cleveland, B.C. Shock, M.W. Cunningham, E.W. Howerth, and M.J. Yabsley. 2018. Necrotizing interstitial

pneumonia and suppurative myocarditis associated with *Bartonella henselae* infection in three Florida pumas. *Journal of Veterinary Diagnostic Investigation* 30(5): 728-732.

- Ferreri, L.L. G. Ortiz, G. Geiger, R. Barriga, R.L. Poulson, A. Gonzalez-Reiche, J. Crum, D.E. Stallknecht, D. Moran, C. Cordon-Rosales, D. Rajao, and D. Perez. 2019. Improved detection of influenza A virus in wild ducks by sequencing directly from swab material. *Ecology and Evolution* doi: 10.1002/ece3.5232.
- Garrett, K., S. Hernandez, G. Balsamo, H. Barron, J. Beasley, J. Brown, E. Cloherty, H. Farid, M. Gabriel, B. Groves, S. Hammer, J. Hill, M. Lewis, K. McManners, N. Nemeth, P. Oesterle, S. Ortiz, L. Peshock, R. Schnellbacher, R. Schott, S. Straif-Bourgeois, and M. Yabsley. Prevalence, distribution, and diversity of cryptic piroplasm infections in raccoons from selected areas of the United States and Canada. *International Journal of Parasitology: Parasites and Wildlife* 9:224-233; doi: 10.1016/j/jippaw.2019.05.007.
- Gettings, J.R., J.E. Lopez, A. Krishnavahjala, B.A. Armstrong, A.T. Thompson, and M.J. Yabsley. 2019. Antibodies to *Borrelia turicatae* in experimentally-infected dogs cross-react with *B. burgdorferi* serologic assays. *Journal of Clinical Biology* pii: JCM 00628-19.
- Gleim, E.R., L.M. Conner, G.E. Zemtsova, M.L. Levin, P. Wong, M.A. Pfaff, and M.J. Yabsley. 2019. Rickettsiales in ticks removed from outdoor workers, Southwest Georgia and Northwest Florida, USA. *Emerging Infectious Diseases* 25(5) 1019-1021.
- Gleim, E.R., G.E. Zemtsova, R.D. Berhaus, M.L. Levin, C. Donner, and M.J. Yabsley. 2019. Frequent prescribed fires can reduce risk of tickborne diseases. *Scientific Reports* 9(1): 9974; doi: 10.1038/s41598-019-46377-4.
- Hollander, L.P., A. Fojtik, C. Kienzle-Dean, N. Davis-Fields, R.L. Poulson, B. Davis, C. Mowry, and D.E. Stallknecht. 2019. Prevalence of influenza A viruses in ducks sampled in northwestern Minnesota and evidence for predominance of H3N8 and H4N6 subtypes in mallards, 2007-2016. Avian Diseases 63 (Suppl) 126-130.
- Kolton, C.B., C.K. Marston, R.A. Stoddard, C.M.
 Cossaboom, J.S. Salzer, T.R. Kozel, M.A. Gates-Hollingsworth, C.A. Cleveland, A.T. Thompson, M.F. Dalton, M.J. Yabsley, and A.R. Hoffmaster.
 2019. Detection of *Bacillus anthracis* in animal tissues using InBios Active Anthrax Detect Rapid

Test Lateral Flow Immunoassay. Letters in Appl. *Microbiology* doi: 10.1111/lam.13134.

- Lewis, J.S., J.L. Corn, J.J. Mayer, T.R. Jordan, M.L. Farnsworth, C.L. Burdett, K.C. VerCauteren, S.J. Sweeney, and R.S. Miller. 2019. Historical, current, and potential population size estimates of invasive wild pigs (*Sus scrofa*) in the United States. *Biological Invasions* 21(7): 2373-2384; doi: 10.1007/s10530-019-01983-1.
- Li., L., A.S. Bowman, T.J. DeLiberto, M.L. Killian, S. Krauss, J.M. Nolting, M.K. Torchetti, A.M. Ramey, A.B. Reeves, D.E. Stallknecht R.J. Webby, and X.F. Wan. 2018. Genetic evidence supports sporadic and independent introductions of subtype H5 low-pathogenic avian influenza A viruses from wild birds to domestic poultry in North America. *Journal of Virology* 92(19). pii: JVI.00913-18.
- Liu, E.W., B.S. Schwartz, N.D. Hysmith, J.P. DeVincenzo, D.T. Larson, R.C. Maves, D.L. Palazzi, C. Meyer, H.T. Cusodio, M.M. Braza, R. Al Hammoud, S. Rao, Y. Qvarnstrom, M.J. Yabsley, R.S. Bradbury, and S.P. Montgomery. 2018. Rat lungworm infection associated with central nervous system disease Eight U.S. states, January 2011-January 2017. *Morbidity and Mortality Weekly Report* 67(30): 825-828.
- Liu, Y., S.K. Nordone, M.J. Yabsley, S.R. Meshnick, R.B. Lund, C.S. McMahan, and J.R. Gettings. 2019. Quantifying the relationship between human Lyme disease and *Borrelia burgdorferi* exposure to domestic dogs. *Geospatial Health* 14(1), doi: 10.4081/gh.2019.750.
- McMillan, J.R., R.A. Blakney, D.G. Mead, S.M. Coker, L.T. Morran, L.A. Waller, U. Kitron, and G.M. Vazquez-Prokopec. 2019. Larviciding *Culex* spp. (Diptera: Culicidae) populations in catch basins and its impact on West Nile virus transmission in urban parks in Atlanta, GA. 2018. *Journal of Medical Entomology* 56(1): 222-232.
- McMillan, J.R., R.A. Blakney, D.G. Mead, W. Koyal, S.M. Coker, L.A. Waller, U. Kitron, and G.M. Vazquez-Prokopec. 2019. Linking transmission potential of multiple vectors to observed patterns of pathogen transmission. *Journal of Applied Ecology* 56: 956-965.
- McMillan, J.R., P.L. Marcet, C.M. Hoover, D.G. Mead, U. Kitron, and G.M. Vazquez-Prokopec. 2019. Host selection of feeding success of *Culex quinquefasciatus* mosquitoes in experimental trials. *Vector-borne and Zoonotic Diseases* https://doi.org/10.1089/ vbz.2018.2381.

- Mendiola, S.Y., M.K. Mills, E. Maki, B.S. Drolet, W.C. Wilson, R. Berghaus, D.E. Stallknecht, J. Breitenbach, D.S. McVey, and M.G. Ruder. 2019. EHDV-2 infection prevalence varies in *Culicoides sonorensis* after feeding on infected white-tailed deer over the course of viremia. *Viruses* 11(4), pii: E371. doi: 10.3390/v11040371
- Mulreany, L.M., K.D. Niedringhaus, H.M.A. Fenton, C. Smith, R. Smith, C.H. Gardiner, B.C. Lewis, T. Craig, B.H. Williams, M.M. Garner, C.A. Cleveland, and M.J. Yabsley. 2018. Filarial dermatitis caused by *Filaria taxidae* in domestic ferrets (*Mustela putorius furo*) from the western United States. *Veterinary Parasitology: Regional Studies and Reports* 14: 155-160.
- Nayduch, D., V. Shankar, J.K. Mills, T. Robl, B.S. Drolet, M.G. Ruder, E.D. Scully, and C.A. Saski. 2019. Transcriptome response of female *Culicoides sonorensis* biting midges (Diptera: Ceratopogonidae) to early infection with epizootic hemorrhagic disease virus (EHDV-2). *Viruses* 11(5); doi: 10.3390/v11050473.
- Niedringhaus, K.D., J.D. Brown, K.M. Sweeley, M.J. Yabsley. 2019. A review of sarcoptic mange in North American wildlife. *International Journal* of *Parasitology: Parasites and Wildlife* 9: 285-297.
- Niedringhaus, K.D., J.D. Brown, M.A. Ternent, W. Childress, J.R. Gettings, M.J. Yabsley. 2019. The emergence and expansion of sarcoptic mange in American black bears (*Ursus americanus*) in the United States. *Veterinary Parasitology: Regional Studies and Reports* doi: 10.1016/j.vprsr.2019.100303.
- Niedringhaus, K.D., H.J. Burchfield, E.J. Elsmo, C.A. Cleveland, H. Fenton, B.C. Shock, C. Muise, J.D. Brown, B. Munk, A. Ellis, R.J. Hall, and M.J. Yabsley. 2019. Trichomonosis due to *Trichomonas gallinae* in barn owls (*Tyto alba*) and barred owls (*Strix varia*) from the eastern United States. *Veterinary Parasitology: Regional Studies* and *Reports* doi: 10.1016/j/vprsr.2019.100281.
- Ramey, A.M., C.A. Cleveland (contributed equally), G.V. Hilderbrand, K. Joly, D. Gustine, B. Mangipane, W.B. Leacock, A.P. Crupi, D.E. Hill, J.P. Dubey, and M.J. Yabsley. 2018. Exposure of Alaska brown bears (*Ursus arctos*) to bacterial, viral, and parasitic agents varies spatiotemporally and may be influenced by age. *Journal of Wildlife Diseases* 55: 576-588.

Ramey, A.M., A.B. Reeves, T. Donnelly, R.L. Poulson, and D.E. Stallknecht. 2018. Introduction of Eurasian-origin influenza A (H8N4) virus into North America by migratory birds. *Emerging Infectious Diseases* 24(10): 1950-1953.

- Ramey, A.M., B.D Uher-Koch, A.B. Reeves, J.A. Schmutz, R.L. Poulson, and D.E. Stallknecht. 2019. Emperor geese (*Anser canagicus*) are exposed to a diversity of influenza A viruses, are infected during the non-breeding period and contribute to intercontinental viral dispersal. *Transboundary and Emerging Diseases* doi: 10.1111/tbed.13226.
- Sapp, S.G.H., S. Handali, S.B. Weinstein, and M.J. Yabsley. 2018. Detection and evaluation of antibody response to a *Baylisascaris*-specific antigen in rodent hosts using Western blotting and ELISA. *Journal of Parasitology* 104(6): 651-659.
- Sapp, S.G.H., M.J. Yabsley, and R.S. Bradbury. 2018. Abnormal helminth egg development, strange morphology, and the identification of intestinal helminth infections. *Emerging Infectious Diseases* 24(8): 1407-1411.
- Schirtzinger, E.E., D.C. Jasperson, M.G. Ruder, D.E. Stallknecht, C. Chase, D.J. Johnson, E.N. Ostlund, and W.C. Wilson. 2019. Evaluation of 2012 US EHDV-2 outbreak isolates from genetic determinants of cattle infection. *Journal of General Virology* 100(4): 556-567.
- Schirtzinger E.E., M.G. Ruder, D.E. Stallknecht, and W.C. Wilson. 2019. Complete genome sequence of a 2016 Bluetongue virus serotype 3 isolate from Louisiana, USA. *Microbiol Resour Announc* 8(19), pii: e00128-19. doi: 10.1128/MRA.00128-19.
- Segovia, K.M., M.S. Franca, C.S. Bahnson, N. Latorre-Margalef, and D.E. Stallknecht. 2018. Are microneutralization and hemagglutination inhibition assays comparable? Serological results from influenza experimentally infected mallard ducks. *Avian Diseases* 63 (Suppl) 138-144.
- Self, S.W., C. McMahan, D.A. Brown, R. Lund, J.R. Gettings, and M.J. Yabsley. 2018. A large scale spatio-temporal binomial regression model for estimating seroprevalence trends. *Environmetrics* 29(8): e2538.
- Self, S.W., C.N. Pulaski, C.S. McMahan, D.A. Brown, M.J. Yabsley, J.R. Gettings. 2019. Regional and local temporal trends in the prevalence of canine heartworm infection in the contiguous United States: 2012–2018. *Parasites* & *Vectors* 12:390. doi: 10.1186/s13071-019-3633-2.

- Sinnott, D., E. Elsmo, S.K. Peltier, M.J. Yabsley, and H. Fenton. 2018. Pathology in practice: Skin lesions in two fox squirrels. *Journal of the American Veterinary Medical Association* 253(6): 723-726.
- Sloyer, K.E, N.D. Burkett-Cadena, A. Yang, J.L. Corn, S.L. Vigil, B.L. McGregor, S.M. Wisely, J.K. Blackburn. 2019. Ecological niche modeling the potential geographic distribution of four *Culicoides* species of veterinary significance in Florida, USA. *PLoS One*: 14(2): e0206648.
- Swanepoel, L., C.A. Cleveland, C. Olfenbuttel, C.G. Dukes, D. Brown, J.D. Brown, A. Surf, R. Tumlison, and M.J. Yabsley. 2018. Prevalence and genetic characterization of *Dirofilaria lutrae* Orihle, 1965 from North American river otters (*Lontra canadensis*). Veterinary Parasitology: Regional Studies and Reports 14: 187-190.
- Swetnam, D., S.G. Widen, T.G. Wood, R. Reyna, L. Wilkerson, M. Debboun, D.A. Symonds, D.G. Mead, B.J. Beaty, H. Guzman, R.B. Tesh, and A.D.T. Barrett. 2018. Terrestrial bird migration and West Nile virus circulation, United States. *Emerging Infectious Diseases* 24(12): 2184-2194.
- Vanstreels, R.E.T., M.J. Yabsley, N.J. Parsons, L. Swanepoel, and P.A. Pistorius. 2018. A novel candidate species of *Anaplasma* that infects avian erythrocytes. *Parasites and Vectors* 11(1): 525.

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